Feb 13, 2017 – 02:19 am GMT

PDB ID : 2XBD
Title : INTERNAL XYLAN BINDING DOMAIN FROM CELLULOMONAS FIMI XYLANASE D, NMR, MINIMIZED AVERAGE STRUCTURE
Authors : Simpson, P.J.; Bolam, D.N.; Cooper, A.; Ciruela, A.; Hazlewood, G.P.; Gilbert, H.J.; Williamson, M.P.
Deposited on : 1998-10-27

This is a Full wwPDB NMR Structure Validation Report for a publicly released PDB entry.

We welcome your comments at validation@mail.wwpdb.org
A user guide is available at http://wwpdb.org/validation/2016/NMRValidationReportHelp
with specific help available everywhere you see the symbol.

The following versions of software and data (see references) were used in the production of this report:

- Cyrange : Kirchner and Güntert (2011)
- NmrClust : Kelley et al. (1996)
- MolProbity : 4.02b-467
- Percentile statistics : 20161228.v01 (using entries in the PDB archive December 28th 2016)
- RCI : v_1n_11_5_13_A (Berjanski et al., 2005)
- PANAV : Wang et al. (2010)
- ShiftChecker : trunk28760
- Ideal geometry (proteins) : Engh & Huber (2001)
- Ideal geometry (DNA, RNA) : Parkinson et al. (1996)
- Validation Pipeline (wwPDB-VP) : recalc28949
1 Overall quality at a glance

The following experimental techniques were used to determine the structure: SOLUTION NMR

The overall completeness of chemical shifts assignment is 57%.

Percentile scores (ranging between 0-100) for global validation metrics of the entry are shown in the following graphic. The table shows the number of entries on which the scores are based.

<table>
<thead>
<tr>
<th>Metric</th>
<th>Whole archive (#Entries)</th>
<th>NMR archive (#Entries)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clashscore</td>
<td>125131</td>
<td>11601</td>
</tr>
<tr>
<td>Ramachandran outliers</td>
<td>121729</td>
<td>10391</td>
</tr>
<tr>
<td>Sidechain outliers</td>
<td>121581</td>
<td>10367</td>
</tr>
</tbody>
</table>

The table below summarises the geometric issues observed across the polymeric chains and their fit to the experimental data. The red, orange, yellow and green segments indicate the fraction of residues that contain outliers for >=3, 2, 1 and 0 types of geometric quality criteria. A cyan segment indicates the fraction of residues that are not part of the well-defined cores, and a grey segment represents the fraction of residues that are not modelled. The numeric value for each fraction is indicated below the corresponding segment, with a dot representing fractions <=5%.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Length</th>
<th>Quality of chain</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>87</td>
<td>55% 40% 5%</td>
</tr>
</tbody>
</table>
2 Ensemble composition and analysis

This entry contains 1 models. Identification of well-defined residues and clustering analysis are not possible.
3  Entry composition

There is only 1 type of molecule in this entry. The entry contains 1192 atoms, of which 577 are hydrogens and 0 are deuteriums.

- Molecule 1 is a protein called XYLANASE D.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Residues</th>
<th>Atoms</th>
<th>Trace</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>87</td>
<td>Total C H N O S</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>1192  368 577 110 134 3</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0</td>
</tr>
</tbody>
</table>
4 Residue-property plots

These plots are provided for all protein, RNA and DNA chains in the entry. The first graphic is the same as shown in the summary in section 1 of this report. The second graphic shows the sequence where residues are colour-coded according to the number of geometric quality criteria for which they contain at least one outlier: green = 0, yellow = 1, orange = 2 and red = 3 or more. Stretches of 2 or more consecutive residues without any outliers are shown as green connectors. Residues which are classified as ill-defined in the NMR ensemble, are shown in cyan with an underline colour-coded according to the previous scheme. Residues which were present in the experimental sample, but not modelled in the final structure are shown in grey.

• Molecule 1: XYLANASE D

Chain A:
5 Refinement protocol and experimental data overview

The models were refined using the following method: *HYBRID DISTANCE GEOMETRY/ SIMULATED ANNEALING*.

Of the 50 calculated structures, 1 were deposited, based on the following criterion: *MINIMIZED AVERAGE STRUCTURE, CALCULATED FROM 38 LOWEST ENERGY STRUCTURES*.

The following table shows the software used for structure solution, optimisation and refinement.

<table>
<thead>
<tr>
<th>Software name</th>
<th>Classification</th>
<th>Version</th>
</tr>
</thead>
<tbody>
<tr>
<td>X-PLOR</td>
<td>refinement</td>
<td>3.1</td>
</tr>
<tr>
<td>MSI FELIX</td>
<td>structure solution</td>
<td>97.0</td>
</tr>
</tbody>
</table>

The following table shows chemical shift validation statistics as aggregates over all chemical shift files. Detailed validation can be found in section 7 of this report.

<table>
<thead>
<tr>
<th>Chemical shift file(s)</th>
<th>BMRB entry 4241</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of chemical shift lists</td>
<td>1</td>
</tr>
<tr>
<td>Total number of shifts</td>
<td>632</td>
</tr>
<tr>
<td>Number of shifts mapped to atoms</td>
<td>632</td>
</tr>
<tr>
<td>Number of unparsed shifts</td>
<td>0</td>
</tr>
<tr>
<td>Number of shifts with mapping errors</td>
<td>0</td>
</tr>
<tr>
<td>Number of shifts with mapping warnings</td>
<td>0</td>
</tr>
<tr>
<td>Assignment completeness (well-defined parts)</td>
<td>57%</td>
</tr>
</tbody>
</table>

No validations of the models with respect to experimental NMR restraints is performed at this time.
6  Model quality

6.1  Standard geometry

There are no covalent bond-length or bond-angle outliers.

Chiral center outliers are detected by calculating the chiral volume of a chiral center and verifying if the center is modelled as a planar moiety or with the opposite hand. A planarity outlier is detected by checking planarity of atoms in a peptide group, atoms in a mainchain group or atoms of a sidechain that are expected to be planar.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Chirality</th>
<th>Planarity</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>All</td>
<td>All</td>
<td>0</td>
<td>3</td>
</tr>
</tbody>
</table>

There are no bond-length outliers.

There are no bond-angle outliers.

There are no chirality outliers.

All planar outliers are listed below.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Res</th>
<th>Type</th>
<th>Group</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>255</td>
<td>ARG</td>
<td>Sidechain</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>302</td>
<td>ARG</td>
<td>Sidechain</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>262</td>
<td>ARG</td>
<td>Sidechain</td>
</tr>
</tbody>
</table>

6.2  Too-close contacts

In the following table, the Non-H and H(model) columns list the number of non-hydrogen atoms and hydrogen atoms in each chain respectively. The H(added) column lists the number of hydrogen atoms added and optimized by MolProbity. The Clashes column lists the number of clashes averaged over the ensemble.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Non-H</th>
<th>H(model)</th>
<th>H(added)</th>
<th>Clashes</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>615</td>
<td>577</td>
<td>574</td>
<td>33</td>
</tr>
<tr>
<td>All</td>
<td>All</td>
<td>615</td>
<td>577</td>
<td>574</td>
<td>33</td>
</tr>
</tbody>
</table>

The all-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The all-atom clashscore for this structure is 28.

All clashes are listed below, sorted by their clash magnitude.

<table>
<thead>
<tr>
<th>Atom-1</th>
<th>Atom-2</th>
<th>Clash(Å)</th>
<th>Distance(Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:A:275:TRP:CZ3</td>
<td>1:A:277:VAL:HG23</td>
<td>0.80</td>
<td>2.11</td>
</tr>
</tbody>
</table>

Continued on next page...
Continued from previous page...

<table>
<thead>
<tr>
<th>Atom-1</th>
<th>Atom-2</th>
<th>Clash(Å)</th>
<th>Distance(Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:A:267:TYR:CE2</td>
<td>1:A:328:ALA:HB2</td>
<td>0.66</td>
<td>2.26</td>
</tr>
<tr>
<td>1:A:275:TRP:CE3</td>
<td>1:A:277:VAL:CG2</td>
<td>0.63</td>
<td>2.80</td>
</tr>
<tr>
<td>1:A:267:VAL:HG11</td>
<td>1:A:267:TYR:CE1</td>
<td>0.63</td>
<td>2.28</td>
</tr>
<tr>
<td>1:A:253:ALA:HB2</td>
<td>1:A:267:TYR:CE2</td>
<td>0.56</td>
<td>2.36</td>
</tr>
<tr>
<td>1:A:267:TYR:CD2</td>
<td>1:A:328:ALA:HB2</td>
<td>0.55</td>
<td>2.36</td>
</tr>
<tr>
<td>1:A:291:TRP:O</td>
<td>1:A:292:ASN:CB</td>
<td>0.52</td>
<td>2.58</td>
</tr>
<tr>
<td>1:A:277:VAL:HG11</td>
<td>1:A:313:PHE:HZ</td>
<td>0.52</td>
<td>1.64</td>
</tr>
<tr>
<td>1:A:265:VAL:CG1</td>
<td>1:A:267:TYR:CE1</td>
<td>0.50</td>
<td>2.94</td>
</tr>
<tr>
<td>1:A:267:TYR:CD2</td>
<td>1:A:328:ALA:CB</td>
<td>0.50</td>
<td>2.95</td>
</tr>
<tr>
<td>1:A:247:THR:CG2</td>
<td>1:A:247:THR:O</td>
<td>0.48</td>
<td>2.60</td>
</tr>
<tr>
<td>1:A:291:TRP:CE2</td>
<td>1:A:314:GLY:HA3</td>
<td>0.48</td>
<td>2.44</td>
</tr>
<tr>
<td>1:A:267:TYR:CE2</td>
<td>1:A:328:ALA:CB</td>
<td>0.48</td>
<td>2.97</td>
</tr>
<tr>
<td>1:A:291:TRP:CE2</td>
<td>1:A:314:GLY:HA3</td>
<td>0.46</td>
<td>2.46</td>
</tr>
<tr>
<td>1:A:289:ALA:HB3</td>
<td>1:A:316:THR:CB</td>
<td>0.46</td>
<td>2.41</td>
</tr>
<tr>
<td>1:A:285:GLN:NE2</td>
<td>1:A:322:SER:HB3</td>
<td>0.45</td>
<td>2.27</td>
</tr>
<tr>
<td>1:A:263:PHE:CZ</td>
<td>1:A:317:VAL:HB</td>
<td>0.44</td>
<td>2.48</td>
</tr>
<tr>
<td>1:A:267:TYR:OH</td>
<td>1:A:326:PRO:O</td>
<td>0.44</td>
<td>2.32</td>
</tr>
<tr>
<td>1:A:253:ALA:HB2</td>
<td>1:A:267:TYR:CD2</td>
<td>0.43</td>
<td>2.49</td>
</tr>
<tr>
<td>1:A:289:ALA:HB3</td>
<td>1:A:316:THR:HB</td>
<td>0.42</td>
<td>1.91</td>
</tr>
<tr>
<td>1:A:277:VAL:HG11</td>
<td>1:A:313:PHE:CZ</td>
<td>0.42</td>
<td>2.48</td>
</tr>
<tr>
<td>1:A:250:SER:O</td>
<td>1:A:269:VAL:HG22</td>
<td>0.41</td>
<td>2.15</td>
</tr>
<tr>
<td>1:A:281:LEU:HD22</td>
<td>1:A:285:GLN:OE1</td>
<td>0.41</td>
<td>2.15</td>
</tr>
<tr>
<td>1:A:292:ASN:HB3</td>
<td>1:A:313:PHE:CB</td>
<td>0.41</td>
<td>2.46</td>
</tr>
<tr>
<td>1:A:289:ALA:HB3</td>
<td>1:A:316:THR:OG1</td>
<td>0.40</td>
<td>2.16</td>
</tr>
<tr>
<td>1:A:292:ASN:HB3</td>
<td>1:A:313:PHE:CA</td>
<td>0.40</td>
<td>2.47</td>
</tr>
</tbody>
</table>

6.3 Torsion angles

6.3.1 Protein backbone

In the following table, the Percentiles column shows the percent Ramachandran outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the backbone conformation was analysed and the total number of residues.
There are no Ramachandran outliers.

### 6.3.2 Protein sidechains

In the following table, the Percentiles column shows the percent sidechain outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the sidechain conformation was analysed and the total number of residues.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Analysed</th>
<th>Rotameric</th>
<th>Outliers</th>
<th>Percentiles</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>68/68 (100%)</td>
<td>53 (78%)</td>
<td>15 (22%)</td>
<td>3, 31</td>
</tr>
<tr>
<td>All</td>
<td>All</td>
<td>68/68 (100%)</td>
<td>53 (78%)</td>
<td>15 (22%)</td>
<td>3, 31</td>
</tr>
</tbody>
</table>

All 15 residues with a non-rotameric sidechain are listed below. They are sorted by the frequency of occurrence in the ensemble.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Res</th>
<th>Type</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>288</td>
<td>GLN</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>292</td>
<td>ASN</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>309</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>324</td>
<td>THR</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>270</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>333</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>325</td>
<td>THR</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>302</td>
<td>ARG</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>298</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>323</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>251</td>
<td>VAL</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>252</td>
<td>THR</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>284</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>250</td>
<td>SER</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>254</td>
<td>THR</td>
</tr>
</tbody>
</table>

### 6.3.3 RNA

There are no RNA molecules in this entry.
6.4 Non-standard residues in protein, DNA, RNA chains

There are no non-standard protein/DNA/RNA residues in this entry.

6.5 Carbohydrates

There are no carbohydrates in this entry.

6.6 Ligand geometry

There are no ligands in this entry.

6.7 Other polymers

There are no such molecules in this entry.

6.8 Polymer linkage issues

There are no chain breaks in this entry.
7 Chemical shift validation

The completeness of assignment taking into account all chemical shift lists is 57% for the well-defined parts and 57% for the entire structure.

7.1 Chemical shift list 1

File name: BMRB entry 4241
Chemical shift list name: assigned_chem_shift_list_1

7.1.1 Bookkeeping

The following table shows the results of parsing the chemical shift list and reports the number of nuclei with statistically unusual chemical shifts.

<table>
<thead>
<tr>
<th>Total number of shifts</th>
<th>632</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of shifts mapped to atoms</td>
<td>632</td>
</tr>
<tr>
<td>Number of unparsed shifts</td>
<td>0</td>
</tr>
<tr>
<td>Number of shifts with mapping errors</td>
<td>0</td>
</tr>
<tr>
<td>Number of shifts with mapping warnings</td>
<td>0</td>
</tr>
<tr>
<td>Number of shift outliers (ShiftChecker)</td>
<td>1</td>
</tr>
</tbody>
</table>

7.1.2 Chemical shift referencing

The following table shows the suggested chemical shift referencing corrections.

<table>
<thead>
<tr>
<th>Nucleus</th>
<th># values</th>
<th>Correction ± precision, ppm</th>
<th>Suggested action</th>
</tr>
</thead>
<tbody>
<tr>
<td>$^{13}$C$_{\alpha}$</td>
<td>0</td>
<td>—</td>
<td>None (insufficient data)</td>
</tr>
<tr>
<td>$^{13}$C$_{\beta}$</td>
<td>0</td>
<td>—</td>
<td>None (insufficient data)</td>
</tr>
<tr>
<td>$^{13}$C$'$</td>
<td>0</td>
<td>—</td>
<td>None (insufficient data)</td>
</tr>
<tr>
<td>$^{15}$N</td>
<td>84</td>
<td>-0.77 ± 0.68</td>
<td>None needed (imprecise)</td>
</tr>
</tbody>
</table>

7.1.3 Completeness of resonance assignments

The following table shows the completeness of the chemical shift assignments for the well-defined regions of the structure. The overall completeness is 57%, i.e. 511 atoms were assigned a chemical shift out of a possible 899. 0 out of 10 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

<table>
<thead>
<tr>
<th></th>
<th>Total</th>
<th>$^1$H</th>
<th>$^{13}$C</th>
<th>$^{15}$N</th>
</tr>
</thead>
<tbody>
<tr>
<td>Backbone</td>
<td>256/431 (59%)</td>
<td>172/172 (100%)</td>
<td>0/174 (0%)</td>
<td>84/85 (99%)</td>
</tr>
<tr>
<td>Sidechain</td>
<td>220/406 (54%)</td>
<td>207/234 (88%)</td>
<td>0/152 (0%)</td>
<td>13/20 (65%)</td>
</tr>
</tbody>
</table>

Continued on next page...
The following table shows the completeness of the chemical shift assignments for the full structure. The overall completeness is 57%, i.e. 511 atoms were assigned a chemical shift out of a possible 899. 0 out of 10 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

<table>
<thead>
<tr>
<th></th>
<th>Total</th>
<th>$^1$H</th>
<th>$^{13}$C</th>
<th>$^{15}$N</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aromatic</td>
<td>35/62 (56%)</td>
<td>32/32 (100%)</td>
<td>0/27 (0%)</td>
<td>3/3 (100%)</td>
</tr>
<tr>
<td>Overall</td>
<td>511/899 (57%)</td>
<td>411/438 (94%)</td>
<td>0/353 (0%)</td>
<td>100/108 (93%)</td>
</tr>
</tbody>
</table>

7.1.4 Statistically unusual chemical shifts

The following table lists the statistically unusual chemical shifts. These are statistical measures, and large deviations from the mean do not necessarily imply incorrect assignments. Molecules containing paramagnetic centres or hemes are expected to give rise to anomalous chemical shifts.

<table>
<thead>
<tr>
<th>Mol</th>
<th>Chain</th>
<th>Res</th>
<th>Type</th>
<th>Atom</th>
<th>Shift, ppm</th>
<th>Expected range, ppm</th>
<th>Z-score</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>267</td>
<td>TYR</td>
<td>HB2</td>
<td>0.87</td>
<td>4.76 - 1.06</td>
<td>-5.5</td>
</tr>
</tbody>
</table>

7.1.5 Random Coil Index (RCI) plots

The image below reports random coil index values for the protein chains in the structure. The height of each bar gives a probability of a given residue to be disordered, as predicted from the available chemical shifts and the amino acid sequence. A value above 0.2 is an indication of significant predicted disorder. The colour of the bar shows whether the residue is in the well-defined core (black) or in the ill-defined residue ranges (cyan), as described in section 2 on ensemble composition.

Random coil index (RCI) for chain A: