

# Integrative Structure Validation Report

October 09, 2025 - 04:40 PM PDT

*The following software was used in the production of this report:*

*IHMValidation Version 3.0*

*Python-IHM Version 2.5*

*EMDB validation analysis Version 0.0.1.dev127*

*ChimeraX Version 1.9*

*Chimera Version 1.19*

*MapQ Version 1.8.1*

PDB ID	9A1P   pdb_00009a1p
PDB-Dev ID	PDBDEV_00000097
Structure Title	Comprehensive structure and functional adaptations of the yeast nuclear pore complex
Structure Authors	Akey CW; Singh D; Ouch C; Echeverria I; Nudelman I; Varberg JM; Yu Z; Fang F; Shi Y; Wang J; Salzberg D; Song K; Xu C; Gumbart JC; Suslov S; Unruh J; Jaspersen SL; Chait BT; Sali A; Fernandez-Martinez J; Ludtke SJ; Villa E; Rout MP
Deposited on	2021-11-19

*This is a PDB-IHM Structure Validation Report.*

*We welcome your comments at [helpdesk@pdb-ihm.org](mailto:helpdesk@pdb-ihm.org)*

*A user guide is available at [https://pdb-ihm.org/validation\\_help.html](https://pdb-ihm.org/validation_help.html) with specific help available everywhere you see the  symbol.*

*List of references used to build this report is available [here](#).*

## 1. Overview

### 1.1. Summary

*This entry consists of 10 model(s). A total of 3 dataset(s) were used to build this entry.*

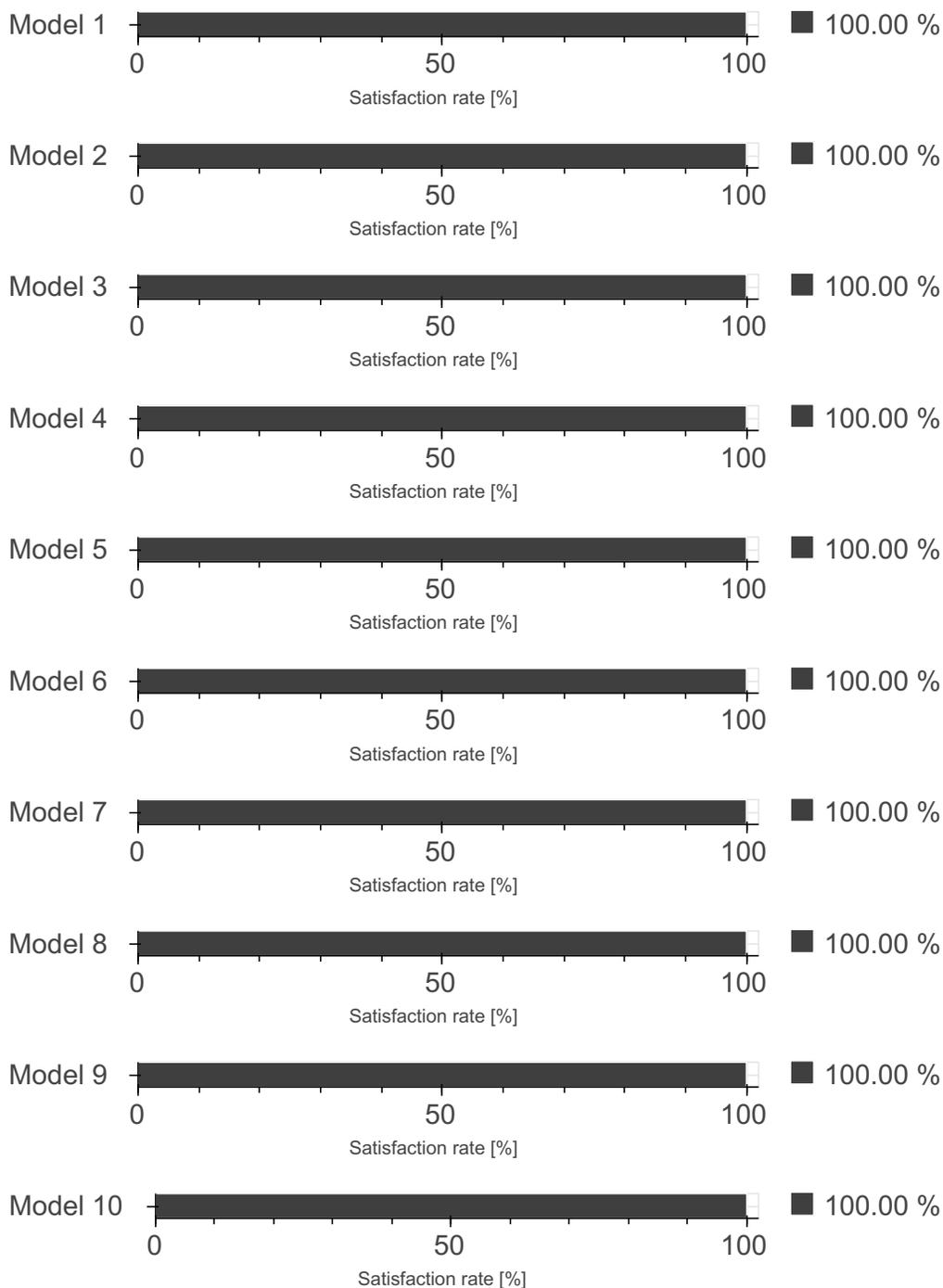
Name	Type	Count
3DEM volume	Experimental data	1

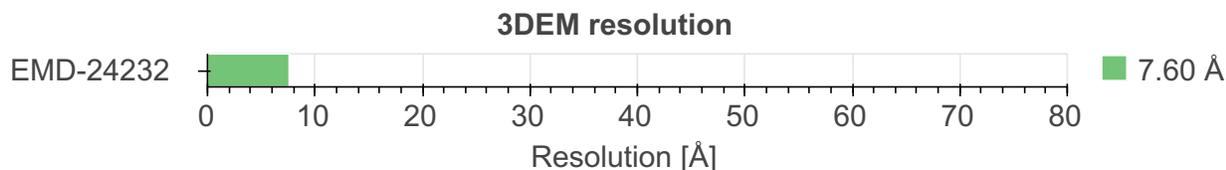
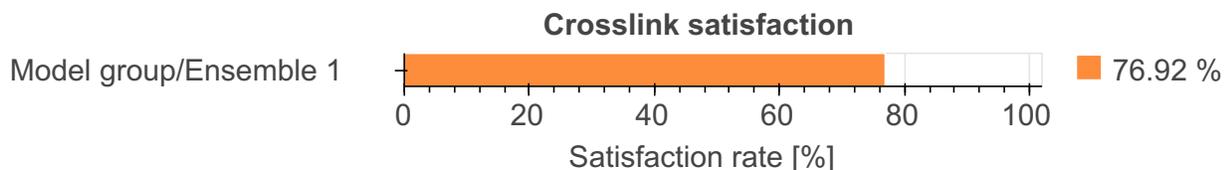
Name	Type	Count
Crosslinking-MS data	Experimental data	1
Experimental model	Starting model	1

## 1.2. Overall quality ?

*This validation report contains model quality assessments for all structures, data quality and fit to model assessments for SAS and crosslinking-MS datasets. Data quality and fit to model assessments for other datasets and model uncertainty are under development. Number of plots is limited to 256.*

### Model Quality: Excluded Volume Analysis ?



Data Quality ?Fit to Data Used for Modeling ?2. Model Details ?2.1. Ensemble information ?

*This entry consists of 1 distinct ensemble(s).*

2.2. Representation ?

*This entry has 1 representation(s).*

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
1	1-10	2	Nup170	0	1502	1-1502	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				Y					
		1	Nup157	1	1391	1-1391	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				Z					
		5	Nsp1	A	823	1-823	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				D					
				G					
				J					
		6	Nup57	B	541	1-541	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				E					
				H					
				K					
		7	Nup49	C	472	1-472	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				F					

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
				I					
				L					
		9	Nup192	M	1683	1-1683	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				O					
		8	Nup188	N	1655	1-1655	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				P					
		10	Nic96	Q	839	1-839	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				R					
				S					
				T					
		3	Nup53	U	475	1-475	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				W					
		4	Nup59	V	528	1-528	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
				X					

### 2.3. Datasets used for modeling

There are 3 unique datasets used to build the models in this entry.

ID	Dataset type	Database name	Data access code
1	Crosslinking-MS data	Zenodo	<a href="https://zenodo.org/record/105281/files/5662389">10.5281/zenodo.5662389</a>
2	3DEM volume	EMDB	<a href="https://www.ebi.ac.uk/emdb/EMD-24232">EMD-24232</a>
3	Experimental model	PDB	<a href="https://www.rcsb.org/structure/pdb_00007n85">pdb_00007n85</a>

### 2.4. Methodology and software

This entry is a result of 1 distinct protocol(s).

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
1	1	Production sampling	Enumeration	Not available	1200	False	True

There are 2 software packages reported in this entry.

ID	Software name	Software version	Software classification	Software location
1	<a href="http://bioinf.cs.ucl.ac.uk/psipred/">PSIPRED</a>	4.00	secondary structure prediction	<a href="http://bioinf.cs.ucl.ac.uk/psipred/">http://bioinf.cs.ucl.ac.uk/psipred/</a>
2	<a href="https://integrativemodeling.org">Integrative Modeling Platform (IMP)</a>	2.20	integrative model building	<a href="https://integrativemodeling.org">https://integrativemodeling.org</a>

### 3. Data quality ?

#### 3.2. Crosslinking-MS

At the moment, data validation is only available for crosslinking-MS data deposited as a fully *compliant* dataset in the *PRIDE Crosslinking* database. Correspondence between crosslinking-MS and entry entities is established using *pyHMMER*. Only residue pairs that passed the reported threshold are used for the analysis. The values in the report have to be interpreted in the context of the experiment (i.e. only a minor fraction of in-situ or in-vivo dataset can be used for modeling).

Crosslinking-MS dataset is not available in the [PRIDE Crosslinking](#) database.

#### 3.3. 3DEM ?

This section describes quality of the 3DEM datasets

##### [EMD-24232](#)

###### 3.3.1. Experimental information ?

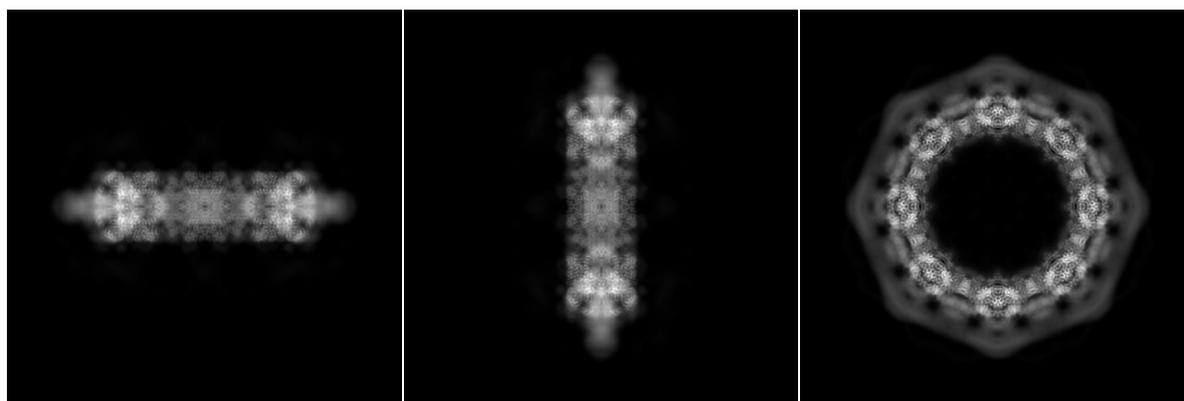
EM reconstruction method:	SINGLE PARTICLE
Resolution:	7.60 Å
Recommended level:	0.595
Estimated volume:	20886.90 nm <sup>3</sup>
Specimen preparation:	Preparation ID 1 Vitrification
Map-only validation report:	<a href="#">wwPDB validation report</a>

###### 3.3.2. Map visualisation ?

This section contains visualisations of the EMDB entry EMD-24232. These allow visual inspection of the internal detail of the map and identification of artifacts. Images derived from a raw map, generated by summing the deposited half-maps, are presented below the corresponding image components of the primary map to allow further visual inspection and comparison with those of the primary map.

###### 3.3.2.1. Orthogonal projections ?

###### Primary map

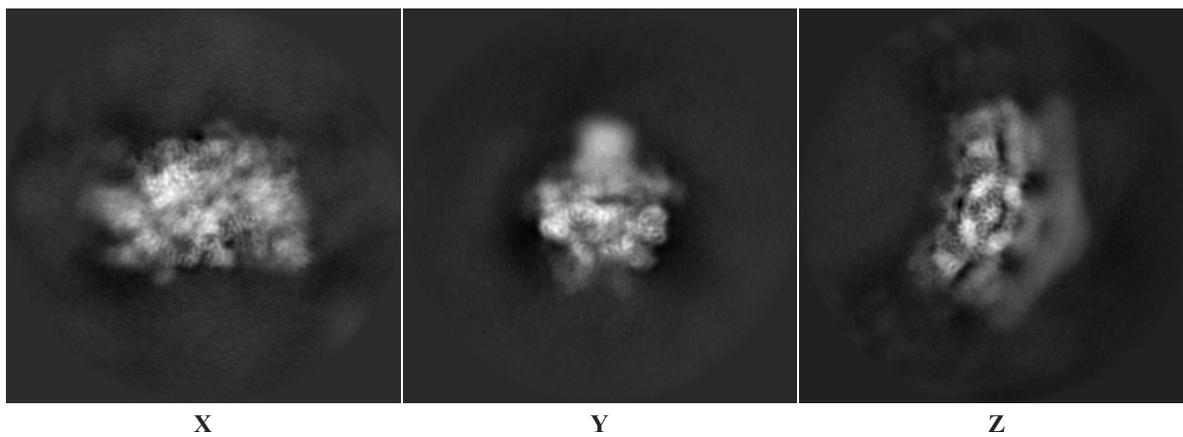


X

Y

Z

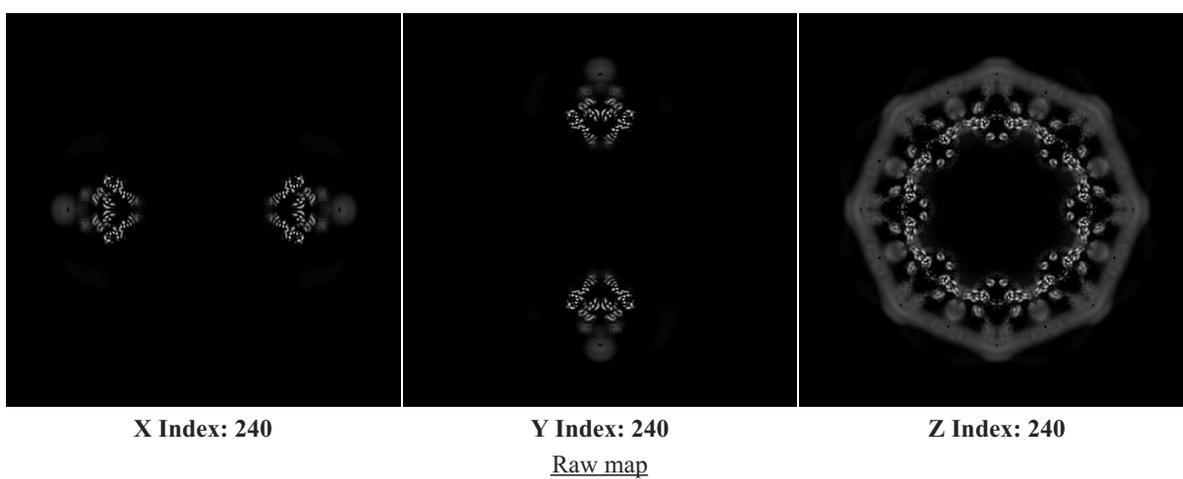
Raw map



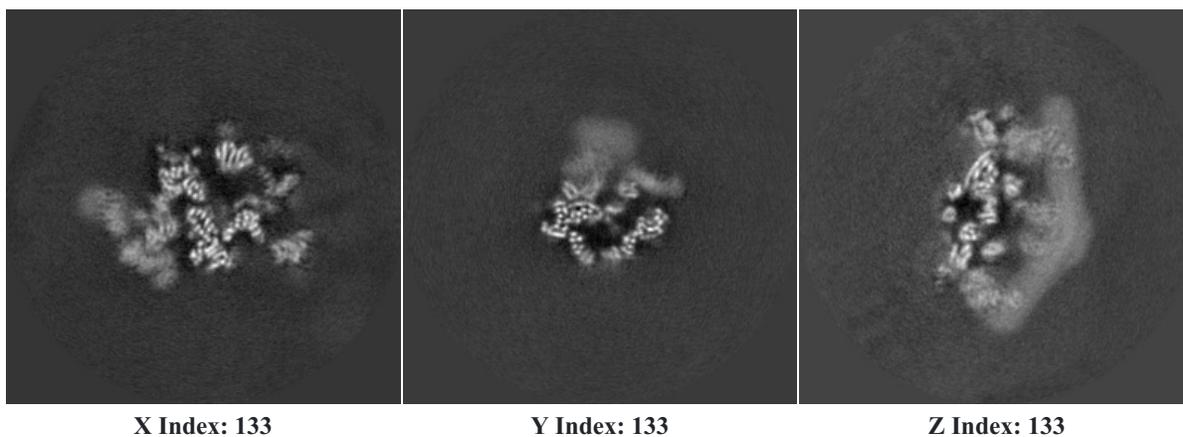
The images above show the map projected in three orthogonal directions.

### 3.3.2.2. Central slices ?

#### Primary map



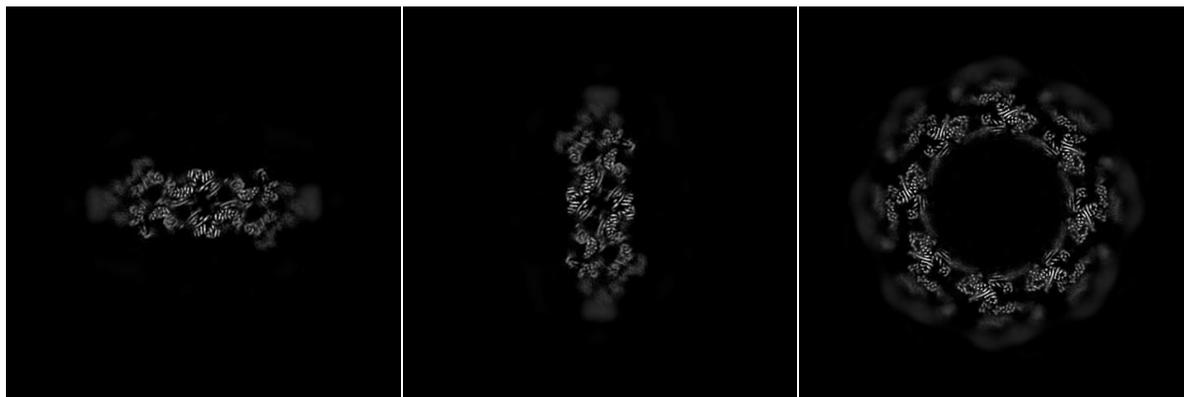
#### Raw map



The images above show central slices of the map in three orthogonal directions.

### 3.3.2.3. Largest variance slices ?

#### Primary map

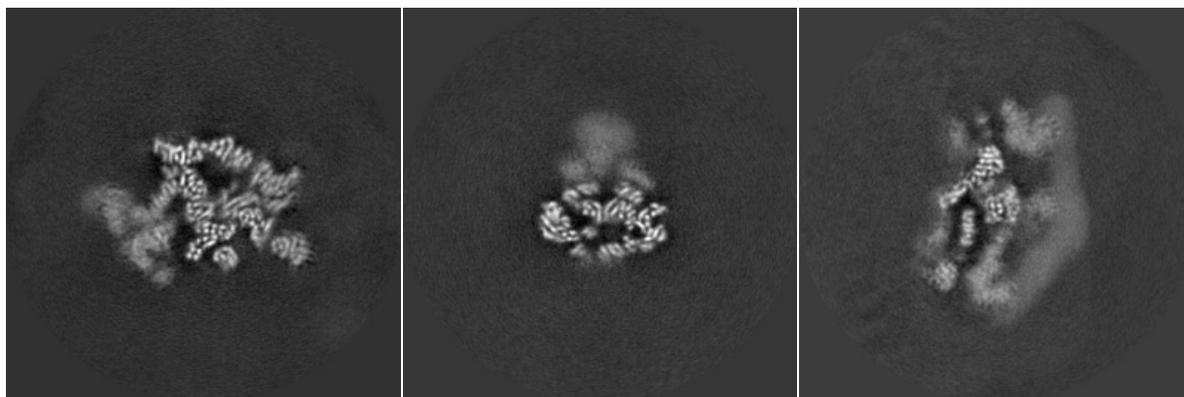


X Index: 340

Y Index: 140

Z Index: 257

Raw map



X Index: 129

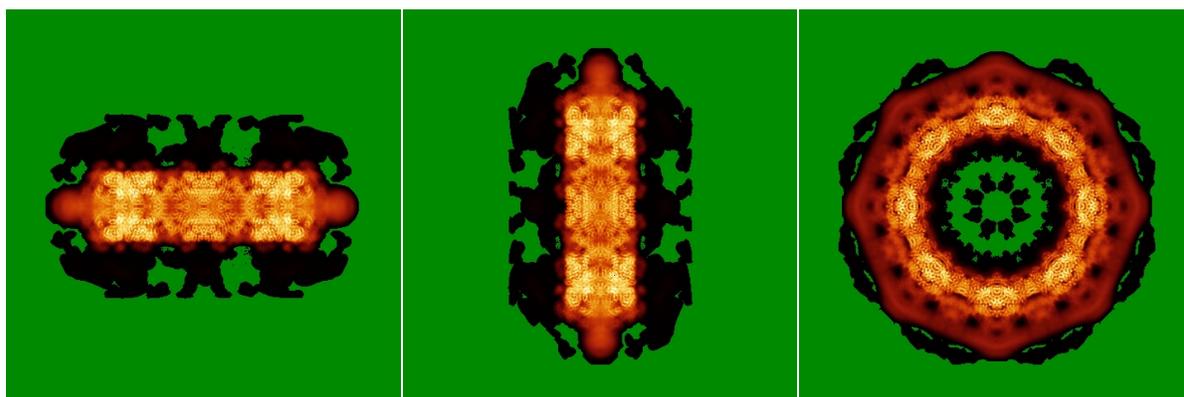
Y Index: 126

Z Index: 125

The images above show the largest variance slices of the map in three orthogonal directions.

#### 3.3.2.4 Orthogonal standard-deviation projections (false-color) ?

Primary map

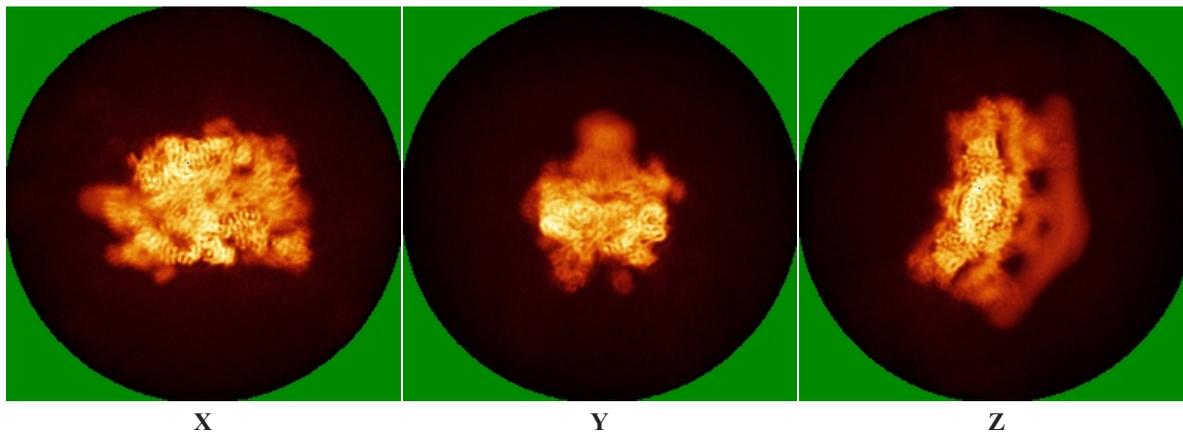


X

Y

Z

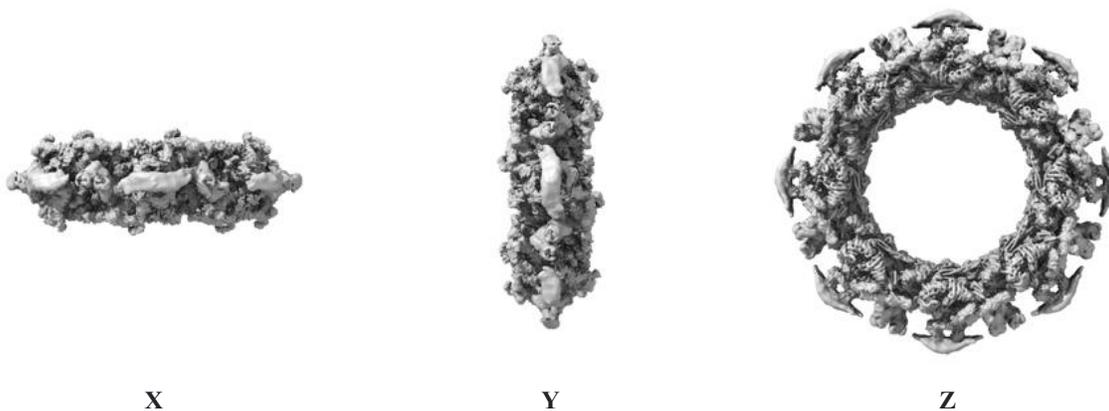
Raw map



The images above show the map standard deviation projections with false color in three orthogonal directions. Minimum values are shown in green, max in blue, and dark to light orange shades represent small to large values respectively.

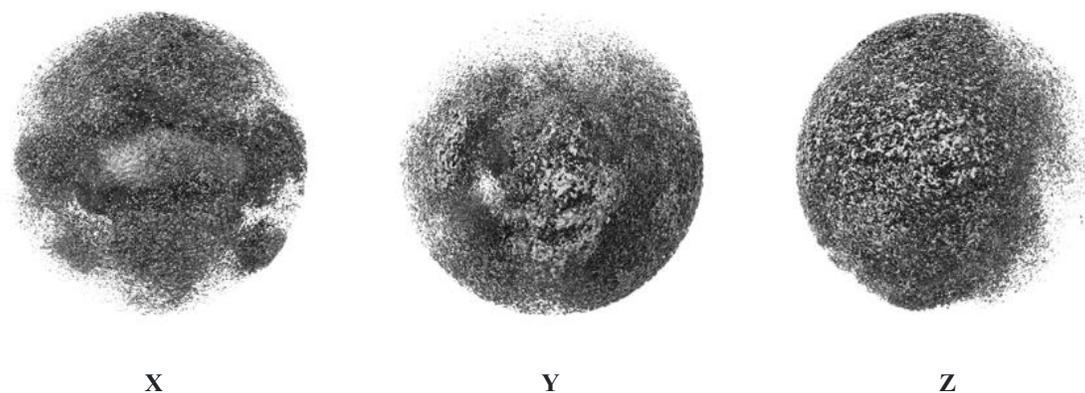
#### 3.3.2.5. Orthogonal surface views [?](#)

##### Primary map



The images above show the 3D surface view of the map at the recommended contour level 0.595 . These images, in conjunction with the slice images, may facilitate assessment of whether an appropriate contour level has been provided.

##### Raw map



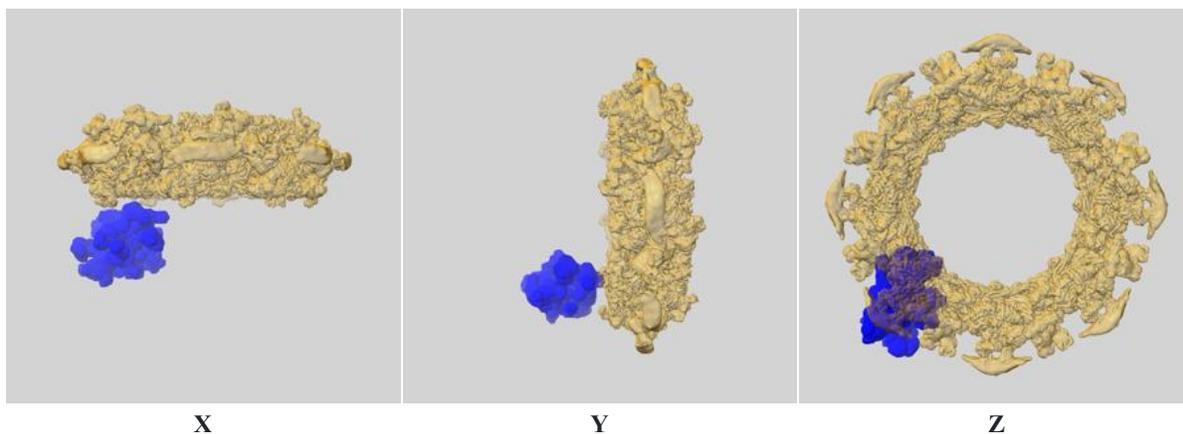
These images show the 3D surface of the raw map. The raw map's contour level 0.090 was selected so that its surface encloses the same volume as the primary map does at its recommended contour level.

#### 3.3.2.6. Mask visualisation [?](#)

This section shows the 3D surface view of the primary map at 50% transparency overlaid with the specified mask at 0% transparency. A mask typically either:

- Encompasses the whole structure;
- Separates out a domain, a functional unit, a monomer or an area of interest from a larger structure.

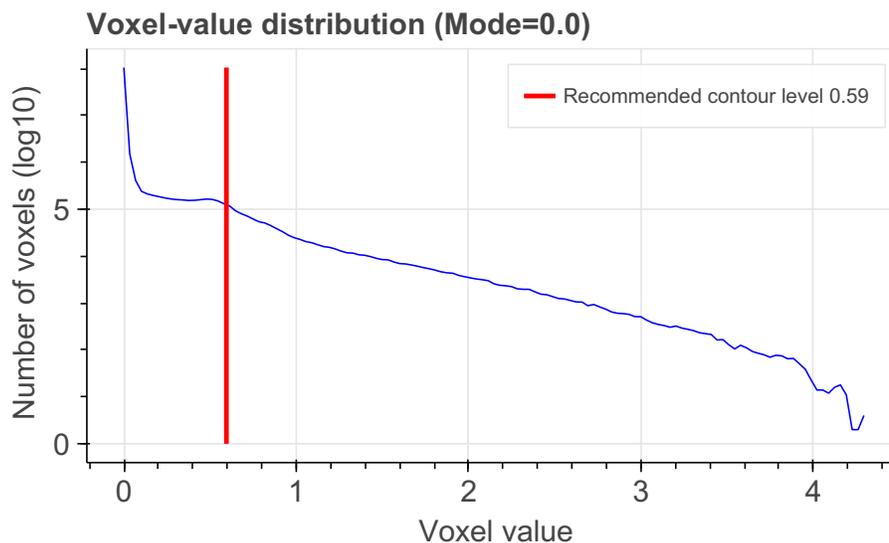
[emd\\_24232\\_msk\\_1.map](#) ?



[3.3.3. Map analysis](#) ?

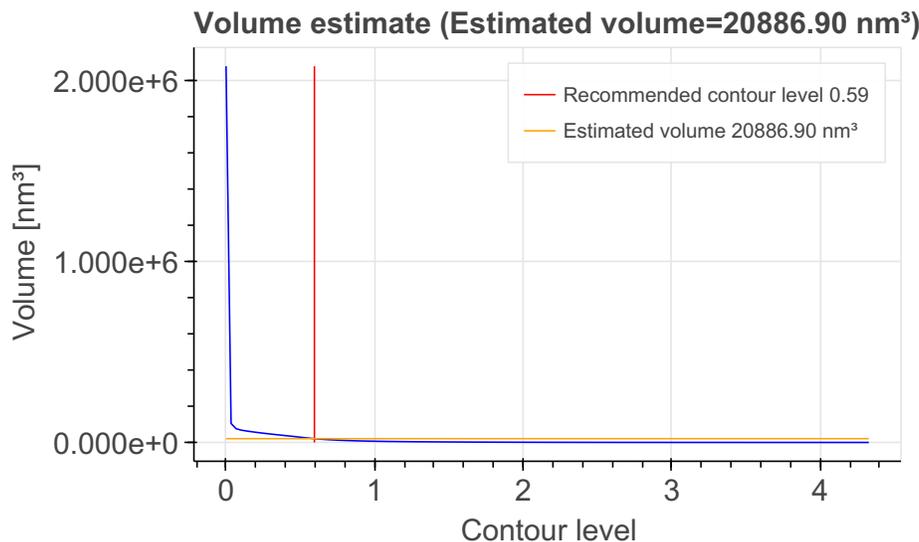
*This section contains the results of statistical analysis of the map.*

[3.3.3.1. Map-value distribution](#) ?



The map-value distribution is plotted in 128 intervals along the x-axis. The y-axis is logarithmic. A spike in this graph at zero usually indicates that the volume has been masked.

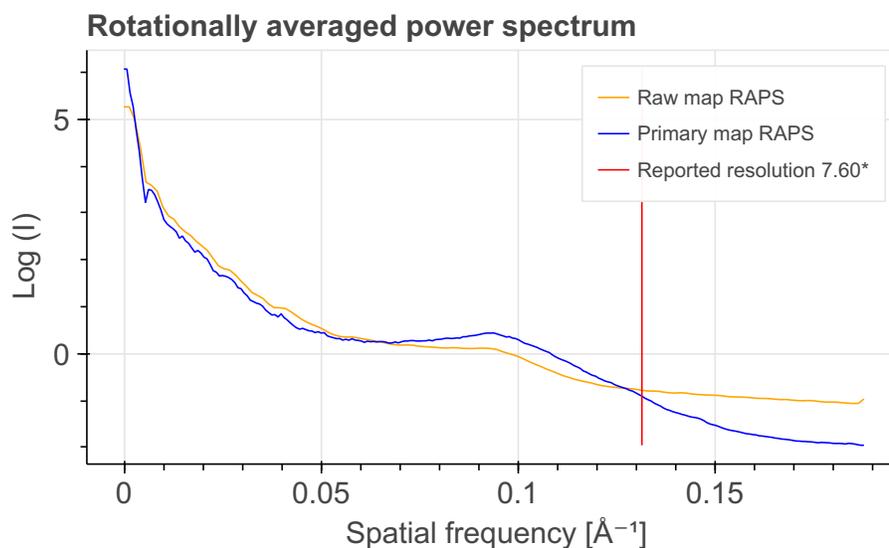
[3.3.3.2. Volume estimate](#) ?



The volume at the recommended contour level is 20886.90 nm<sup>3</sup>.

The volume estimate graph shows how the enclosed volume varies with the contour level. The recommended contour level is shown as a vertical line and the intersection between the line and the curve gives the volume of the enclosed surface at the given level.

### 3.3.3.3. Rotationally averaged power spectrum

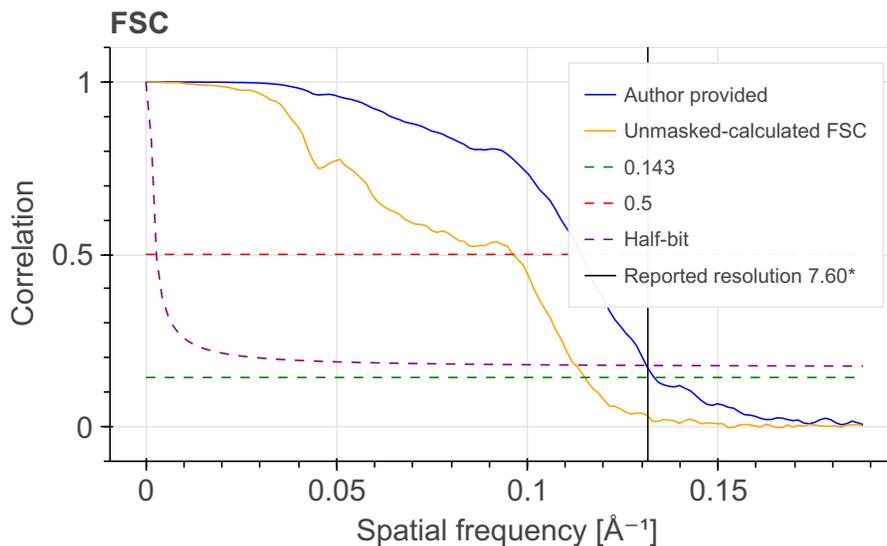


\*Reported resolution corresponds to spatial frequency of 0.132 Å<sup>-1</sup>

### 3.3.4. Fourier-Shell correlation

#### 3.3.4.1. FSC

Fourier-Shell Correlation (FSC) is the most commonly used method to estimate the resolution of single-particle and subtomogram-averaged maps. The shape of the curve depends on the imposed symmetry, mask and whether or not the two 3D reconstructions used were processed from a common reference. The reported resolution is shown as a black line. A curve is displayed for the half-bit criterion in addition to lines showing the 0.143 gold standard cut-off and 0.5 cut-off.



\*Reported resolution corresponds to spatial frequency of  $0.132 \text{ \AA}^{-1}$

### 3.3.4.2. Resolution estimates ?

Resolution estimate ( $\text{\AA}$ )	Estimation criterion (FSC cut-off)		
	0.143	0.5	Half-bit
Reported by author	7.60	-	-
Author-provided FSC curve	7.51	8.76	7.62
Unmasked-calculated*	8.69	10.40	8.88

\*Resolution estimate based on FSC curve calculated by comparison of deposited half-maps. The value from deposited half-maps intersecting FSC 0.143 CUT-OFF 8.69 differs from the reported value 7.60 by more than 10%.

## 4. Model quality ?

For models with atomic structures, MolProbity analysis is performed. For models with coarse-grained or multi-scale structures, excluded volume analysis is performed.

### 4.1a. Excluded Volume Analysis ?

Excluded volume satisfaction for the models in the entry are listed below. The Analysed column shows the number of particle-particle or particle-atom pairs for which excluded volume was analysed.

Model ID	Analysed	Number of violations	Excluded Volume Satisfaction (%)
1	7541209455	124879	100.00
2	7541209455	124879	100.00
3	7541209455	124879	100.00
4	7541209455	124879	100.00
5	7541209455	124879	100.00
6	7541209455	124879	100.00
7	7541209455	124879	100.00

Model ID	Analysed	Number of violations	Excluded Volume Satisfaction (%)
8	7541209455	124879	100.00
9	7541209455	124879	100.00
10	7541209455	124879	100.00

## 5. Fit to Data Used for Modeling Assessment ?

### 5.2. Crosslinking-MS ?

#### 5.2.1. Restraint types ?

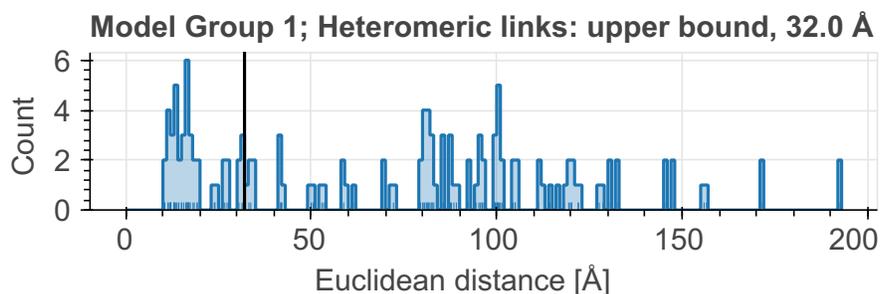
This table summarizes information about crosslinker(s) used for data generation, and how crosslinking information was translated into actual modeling restraints. Restraints assigned "by-residue" are interpreted as between CA atoms. Restraints between coarse-grained beads are indicated as "coarse-grained". *Restraint group* represents a set of crosslinking restraints applied collectively in the modeling.

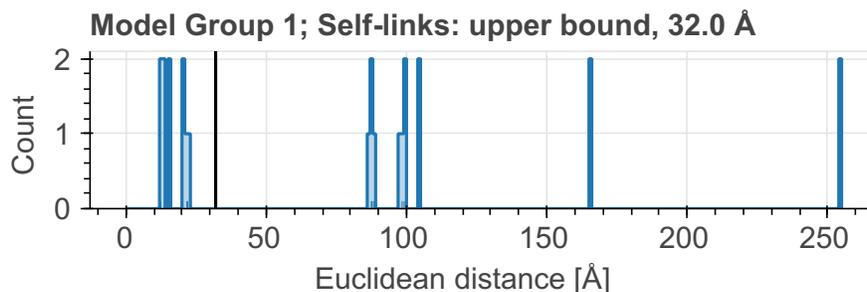
There are 1284 crosslinking restraints combined in 114 restraint groups.

Linker	Residue 1	Atom 1	Residue 2	Atom 2	Restraint type	Distance, Å	Count
DSS	ALA	CA	LYS	CA	upper bound	32.00	692
DSS	LYS	CA	LYS	CA	upper bound	32.00	512
DSS	ALA	CA	ALA	CA	upper bound	32.00	80

#### Distograms of individual restraints

Distograms (i.e., histogram plots of distances) provide an overview of distributions of distances between residues for which chemical crosslinks were identified. The shift of the distogram relative to the threshold value may indicate a poor model. Restraints with identical thresholds are grouped into one plot. Only the best distance per restraint per model group/ensemble is plotted. Inter- and intramolecular (including self-links) restraints are also grouped into one plot. Distance for a restraint between coarse-grained beads is calculated as a minimal distance between shells; if beads intersect, the distance will be reported as 0.0. A bead with the highest available resolution for a given residue is used for the assessment.





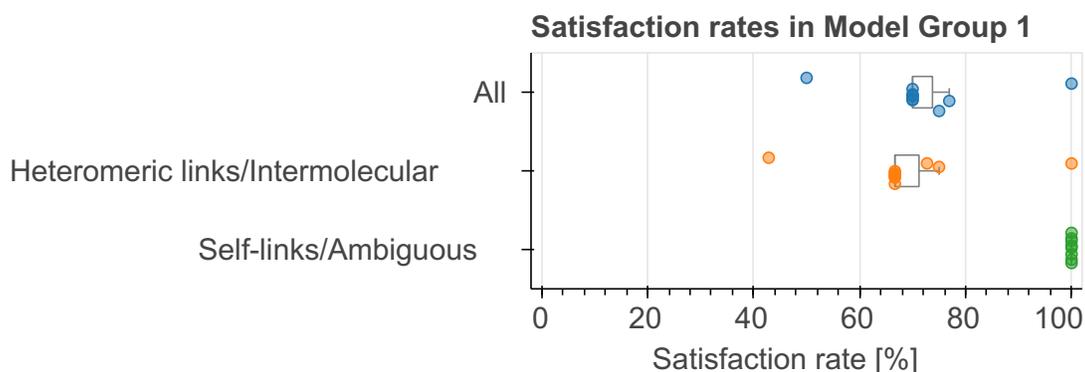
### 5.2.2. Satisfaction of restraints ?

Satisfaction of restraints is calculated on a *restraint group* (a set of crosslinking restraints applied collectively in the modeling) level. Satisfaction of a restraint group depends on satisfaction of individual restraints in the group and the conditionality (all/any). A restraint group is considered satisfied, if the condition was met in at least one model of the model group/ensemble. The number of measured restraints can be smaller than the total number of restraint groups if crosslinks involve non-modeled residues. Only deposited models are used for validation right now.

State group	State	Model group	# of Deposited models/Total	Restraint group type	Satisfied (%)	Violated (%)	Count (Total=114)
1	1	1	10/10	All	76.92	23.08	13
				Heteromeric links/ Intermolecular	72.73	27.27	11
				Self-links/ Ambiguous	100.00	0.00	2

#### Per-model satisfaction rates in ensembles

Every point represents one model in a model group/ensemble. Where possible, boxplots with quartile marks are also plotted.



### 5.3. 3DEM

This section describes fit of models to the 3DEM data. Only results for the representative model, selected as a first model with the largest number of asymmetric units.

3DEM validation for coarse-grained structures is under development.

## 6. Fit to Data Used for Validation Assessment ?

Validation for this section is under development.

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### *Acknowledgments*

*The development of integrative model validation metrics, implementation of a model validation pipeline, and creation of a validation report for integrative structures are funded by NSF awards to the [PDB-IHM team](#) (DBI-1756248, DBI-2112966, DBI-2112967, DBI-2112968, and DBI-1756250) and awards from NSF, NIH, and DOE to the [RCSB PDB](#) (DBI-2321666, R01GM157729, and DE-SC0019749). The PDB-IHM team and members of the [Sali lab](#) contributed model validation metrics and software packages.*

*Dr. Jill Trewhella, Dr. Dina Schneidman, and members of the [SASBDB](#) repository are acknowledged for their advice and support in implementing SAS validation methods. Team members from the labs of Dr. Juri Rappsilber, Dr. Alexander Leitner, Dr. Andrea Graziadei, and members of [PRIDE](#) database are acknowledged for their advice and support in implementing crosslinking-MS validation methods. We are grateful to Dr. Shruthi Viswanath for discussions about uncertainty assessment of integrative structural models.*

*Members of the [wwPDB Integrative/Hybrid Methods Task Force](#) provided recommendations and community support for the project.*