

# Integrative Structure Validation Report

October 09, 2025 - 04:41 PM PDT

*The following software was used in the production of this report:*

*IHMValidation Version 3.0*

*Python-IHM Version 2.5*

*MolProbity Version 4.5.2*

*EMDB validation analysis Version 0.0.1.dev127*

*ChimeraX Version 1.9*

*Chimera Version 1.19*

*MapQ Version 1.8.1*

PDB ID	9A2D   pdb_00009a2d
PDB-Dev ID	PDBDEV_00000156
Structure Title	Modeling of the ciliary Intraflagellar transport-A complex
Structure Authors	McCafferty, C.L.; Papoulas, O.; Jordan, M.A.; Hoogerbrugge, G.; Nichols, C.; Pigino, G.; Taylor, D.W.; Wallingford, J.B.; Marcotte, E.M.
Deposited on	2022-08-03

*This is a PDB-IHM Structure Validation Report.*

*We welcome your comments at [helpdesk@pdb-ihm.org](mailto:helpdesk@pdb-ihm.org)*

*A user guide is available at [https://pdb-ihm.org/validation\\_help.html](https://pdb-ihm.org/validation_help.html) with specific help available everywhere you see the  symbol.*

*List of references used to build this report is available [here](#).*

## 1. Overview

### 1.1. Summary

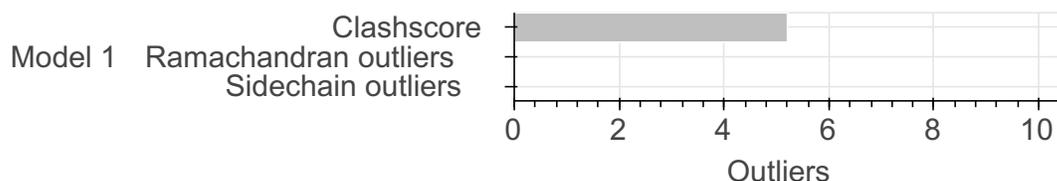
*This entry consists of 1 model(s). A total of 3 dataset(s) were used to build this entry.*

Name	Type	Count
3DEM volume	Experimental data	1
Crosslinking-MS data	Experimental data	1
De Novo model	Starting model	1

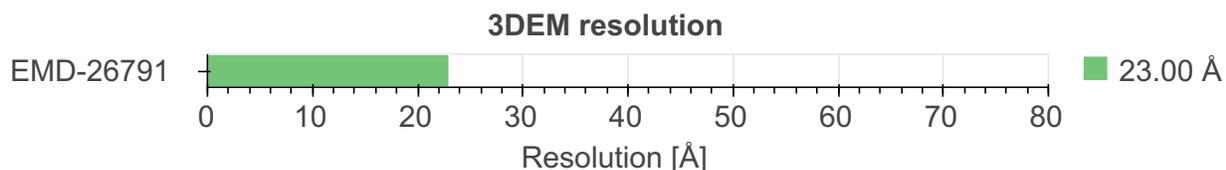
## 1.2. Overall quality [?](#)

This validation report contains model quality assessments for all structures, data quality and fit to model assessments for SAS and crosslinking-MS datasets. Data quality and fit to model assessments for other datasets and model uncertainty are under development. Number of plots is limited to 256.

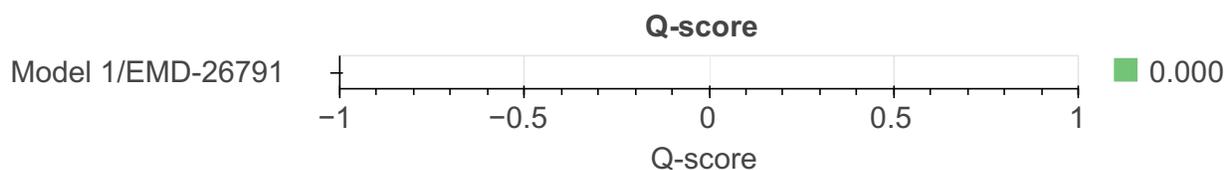
### Model Quality: MolProbity Analysis [?](#)



### Data Quality [?](#)



### Fit to Data Used for Modeling [?](#)



## 2. Model Details [?](#)

### 2.1. Ensemble information [?](#)

This entry consists of 0 distinct ensemble(s).

### 2.2. Representation [?](#)

This entry has 1 representation(s).

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
1	1	1	Intraflagellar transport protein 43	A	146	70-80, 90-130	1-69, 81-89, 131-146	100.00 / 35.62	Coarse-grained: 1 residue(s) per bead
		2	Intraflagellar transport protein 121	B	1195	1-341, 342-655, 656-799, 800-895, 896-978, 979-1004, 1005-1195	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
		3	Intraflagellar transport protein 122	C	1251	1-319, 320-581, 582-811, 812-1251	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
		4	Intraflagellar transport protein 139	D	1334	1-1334	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
		5	Intraflagellar transport protein 140	E	1407	1-376, 377-713, 714-979, 980-1080, 1081-1407	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead
		6	Intraflagellar transport protein 144	F	1387	1-655, 656-985, 986-1114, 1115-1387	-	100.00 / 100.00	Coarse-grained: 1 residue(s) per bead

### 2.3. Datasets used for modeling

There are 3 unique datasets used to build the models in this entry.

ID	Dataset type	Database name	Data access code
1	De Novo model	Zenodo	<a href="https://zenodo.org/record/7222413">10.5281/zenodo.7222413</a>
2	Crosslinking-MS data	Zenodo	<a href="https://zenodo.org/record/7222413">10.5281/zenodo.7222413</a>
3	3DEM volume	EMDB	<a href="https://www.ebi.ac.uk/emdb/EMD-26791">EMD-26791</a>

### 2.4. Methodology and software

This entry is a result of 1 distinct protocol(s).

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
1	1	Sampling	Replica exchange monte carlo	Not available	200000	False	True

There are 3 software packages reported in this entry.

ID	Software name	Software version	Software classification	Software location
3	<a href="https://alphafold.ebi.ac.uk/">AlphaFold2</a>	Not available	structure prediction	<a href="https://alphafold.ebi.ac.uk/">https://alphafold.ebi.ac.uk/</a>
1	<a href="https://integrativemodeling.org">IMP PMI module</a>	2.11.1	integrative model building	<a href="https://integrativemodeling.org">https://integrativemodeling.org</a>

ID	Software name	Software version	Software classification	Software location
2	<a href="#">Integrative Modeling Platform (IMP)</a>	2.11.1	integrative model building	<a href="https://integrativemodeling.org">https://integrativemodeling.org</a>

### 3. Data quality

#### 3.2. Crosslinking-MS

At the moment, data validation is only available for crosslinking-MS data deposited as a fully *compliant* dataset in the *PRIDE Crosslinking* database. Correspondence between crosslinking-MS and entry entities is established using *pyHMMER*. Only residue pairs that passed the reported threshold are used for the analysis. The values in the report have to be interpreted in the context of the experiment (i.e. only a minor fraction of in-situ or in-vivo dataset can be used for modeling).

Crosslinking-MS dataset is not available in the [PRIDE Crosslinking](#) database.

#### 3.3. 3DEM

This section describes quality of the 3DEM datasets

##### [EMD-26791](#)

###### 3.3.1. Experimental information

EM reconstruction method:	SUBTOMOGRAM AVERAGING
Resolution:	23.00 Å
Recommended level:	1.150
Estimated volume:	1351.55 nm <sup>3</sup>
Specimen preparation:	Preparation ID 1 Vitrification
Map-only validation report:	<a href="#">wwPDB validation report</a>

###### 3.3.2. Map visualisation

This section contains visualisations of the EMDB entry EMD-26791. These allow visual inspection of the internal detail of the map and identification of artifacts. Images derived from a raw map, generated by summing the deposited half-maps, are presented below the corresponding image components of the primary map to allow further visual inspection and comparison with those of the primary map.

###### 3.3.2.1. Orthogonal projections

###### [Primary map](#)



X

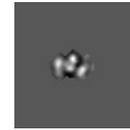
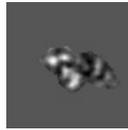
Y

Z

The images above show the map projected in three orthogonal directions.

### 3.3.2.2. Central slices ?

Primary map



X Index: 40

Y Index: 40

Z Index: 40

The images above show central slices of the map in three orthogonal directions.

### 3.3.2.3. Largest variance slices ?

Primary map



X Index: 36

Y Index: 37

Z Index: 30

The images above show the largest variance slices of the map in three orthogonal directions.

### 3.3.2.4 Orthogonal standard-deviation projections (false-color) ?

Primary map



X



Y



Z

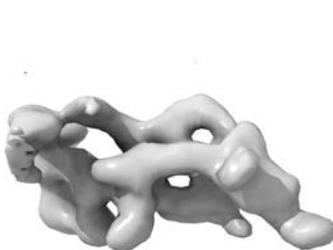
The images above show the map standard deviation projections with false color in three orthogonal directions. Minimum values are shown in green, max in blue, and dark to light orange shades represent small to large values respectively.

### 3.3.2.5. Orthogonal surface views [?](#)

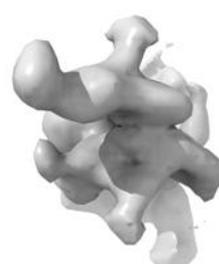
#### Primary map



X



Y



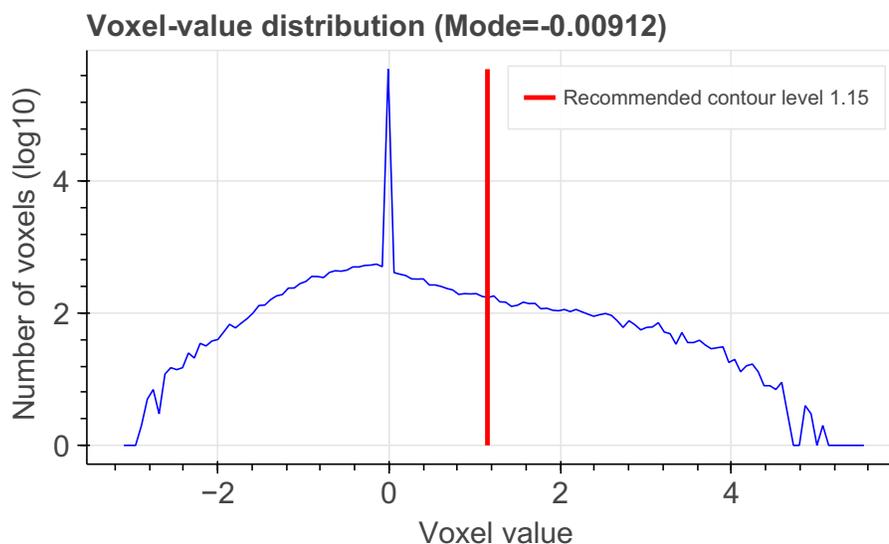
Z

The images above show the 3D surface view of the map at the recommended contour level 1.150. These images, in conjunction with the slice images, may facilitate assessment of whether an appropriate contour level has been provided.

### 3.3.3. Map analysis [?](#)

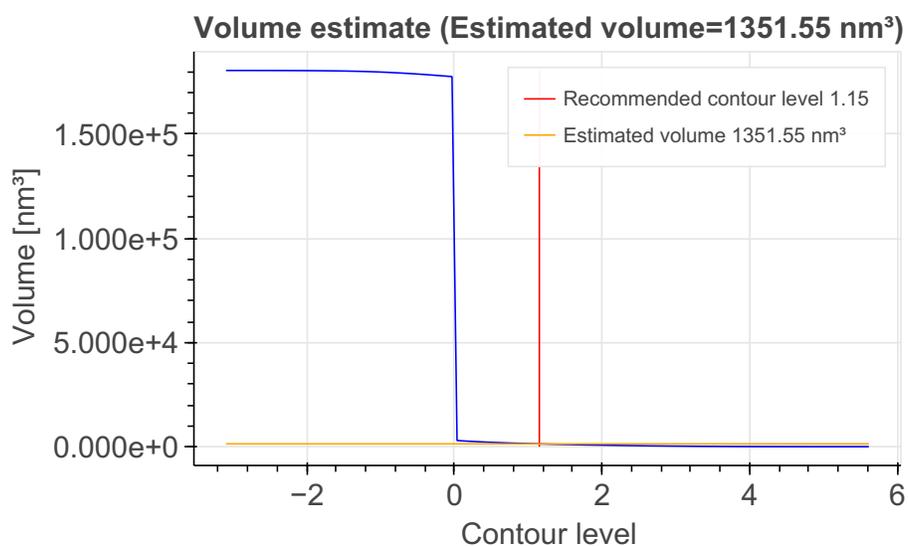
*This section contains the results of statistical analysis of the map.*

#### 3.3.3.1. Map-value distribution [?](#)



The map-value distribution is plotted in 128 intervals along the x-axis. The y-axis is logarithmic. A spike in this graph at zero usually indicates that the volume has been masked.

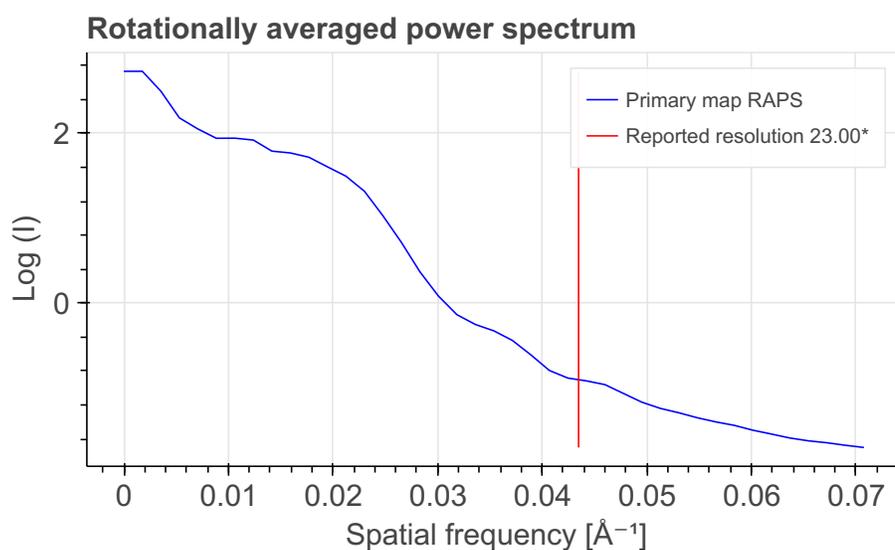
## 3.3.3.2. Volume estimate ?



The volume at the recommended contour level is 1351.55 nm<sup>3</sup>.

The volume estimate graph shows how the enclosed volume varies with the contour level. The recommended contour level is shown as a vertical line and the intersection between the line and the curve gives the volume of the enclosed surface at the given level.

## 3.3.3.3. Rotationally averaged power spectrum ?



\*Reported resolution corresponds to spatial frequency of 0.043 Å<sup>-1</sup>

## 3.3.4. Fourier-Shell correlation ?

## 3.3.4.2. Resolution estimates ?

Resolution estimate (Å)	Estimation criterion (FSC cut-off)		
	0.143	0.5	Half-bit
Reported by author	-	23.00	-

Author-provided FSC curve is not available.

## 4. Model quality ?

For models with atomic structures, MolProbity analysis is performed. For models with coarse-grained or multi-scale structures, excluded volume analysis is performed.

#### 4.1b. MolProbity Analysis

Excluded volume satisfaction for the models in the entry are listed below. The Analysed column shows the number of particle-particle or particle-atom pairs for which excluded volume was analysed.

##### Standard geometry: bond outliers

Bond length outliers can not be evaluated for this model

##### Standard geometry: angle outliers

Bond angle outliers can not be evaluated for this model

##### Too-close contacts

The following all-atom clashscore is based on a MolProbity analysis. All-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The table below contains clashscores for all atomic models in this entry.

Model ID	Clash score	Number of clashes
1	5.21	35

There are 35 clashes. The table below contains the detailed list of all clashes based on a MolProbity analysis. Bad clashes are  $\geq 0.4$  Angstrom.

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
A:102:PRO:CA	D:1048:ASN:CA	1.47	1	1
E:51:TYR:CA	F:1067:ALA:CA	1.43	1	1
A:48:GLN:CA	A:49:GLU:CA	1.34	1	1
F:290:ILE:CA	F:630:ARG:CA	1.22	1	1
C:905:LEU:CA	E:1081:LYS:CA	1.13	1	1
C:710:PRO:CA	E:1369:LEU:CA	1.11	1	1
C:899:MET:CA	E:1078:GLN:CA	1.11	1	1
C:682:ASN:CA	E:1376:GLU:CA	1.09	1	1
D:862:GLY:CA	D:954:GLU:CA	1.05	1	1
C:706:THR:CA	E:1393:ASN:CA	1.03	1	1
C:901:ALA:CA	E:1079:ARG:CA	0.92	1	1
F:764:GLU:CA	F:791:LEU:CA	0.83	1	1
A:132:GLY:CA	A:133:ASN:CA	0.82	1	1
C:1240:GLU:CA	E:1045:LYS:CA	0.76	1	1
C:1190:CYS:CA	E:1083:ARG:CA	0.74	1	1
C:702:ARG:CA	E:1394:LYS:CA	0.72	1	1
C:902:THR:CA	E:1079:ARG:CA	0.69	1	1
F:786:LEU:CA	F:808:ALA:CA	0.68	1	1

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
C:712:ASN:CA	E:1372:TYR:CA	0.66	1	1
F:785:TYR:CA	F:806:ARG:CA	0.63	1	1
F:787:THR:CA	F:806:ARG:CA	0.59	1	1
C:901:ALA:CA	E:1080:ASN:CA	0.58	1	1
F:290:ILE:CA	F:631:GLY:CA	0.51	1	1
C:902:THR:CA	E:1076:GLU:CA	0.49	1	1
A:5:GLY:CA	A:9:TRP:CA	0.49	1	1
F:786:LEU:CA	F:805:ARG:CA	0.49	1	1
C:902:THR:CA	E:1078:GLN:CA	0.48	1	1
F:600:ILE:CA	F:716:ASP:CA	0.48	1	1
F:618:LYS:CA	F:703:LEU:CA	0.48	1	1
C:910:ASN:CA	E:1084:ASP:CA	0.47	1	1
F:787:THR:CA	F:807:LEU:CA	0.47	1	1
F:479:PHE:CA	F:480:PRO:CA	0.41	1	1
E:52:GLU:CA	F:1068:LYS:CA	0.41	1	1
C:1192:THR:CA	E:1084:ASP:CA	0.41	1	1
C:705:ASP:CA	E:1394:LYS:CA	0.40	1	1

### Torsion angles: Protein backbone ?

In the following table, Ramachandran outliers are listed. The Analysed column shows the number of residues for which the backbone conformation was analysed.

Model ID	Analysed	Favored	Allowed	Outliers
1	0	0	0	0

### Torsion angles : Protein sidechains ?

In the following table, sidechain rotameric outliers are listed. The Analysed column shows the number of residues for which the sidechain conformation was analysed.

Model ID	Analysed	Favored	Allowed	Outliers
1	0	0	0	0

## 5. Fit to Data Used for Modeling Assessment ?

### 5.2. Crosslinking-MS ?

#### 5.2.1. Restraint types ?

This table summarizes information about crosslinker(s) used for data generation, and how crosslinking information was translated into actual modeling restraints. Restraints assigned "by-residue" are interpreted as between CA atoms. Restraints between coarse-grained beads are indicated as "coarse-grained". *Restraint group* represents a set of crosslinking restraints applied collectively in the

modeling.

There are 100 crosslinking restraints combined in 100 restraint groups.

Linker	Residue 1	Atom 1	Residue 2	Atom 2	Restraint type	Distance, Å	Count
DSSO	LYS	CA	LYS	CA	upper bound	21.00	96
DSSO	GLU	CA	GLU	CA	upper bound	21.00	2
DSSO	ASP	CA	GLU	CA	upper bound	21.00	1
DSSO	LYS	CA	TYR	CA	upper bound	21.00	1

#### Distograms of individual restraints

Distograms (i.e., histogram plots of distances) provide an overview of distributions of distances between residues for which chemical crosslinks were identified. The shift of the distogram relative to the threshold value may indicate a poor model. Restraints with identical thresholds are grouped into one plot. Only the best distance per restraint per model group/ensemble is plotted. Inter- and intramolecular (including self-links) restraints are also grouped into one plot. Distance for a restraint between coarse-grained beads is calculated as a minimal distance between shells; if beads intersect, the distance will be reported as 0.0. A bead with the highest available resolution for a given residue is used for the assessment.

#### 5.2.2. Satisfaction of restraints

Satisfaction of restraints is calculated on a *restraint group* (a set of crosslinking restraints applied collectively in the modeling) level. Satisfaction of a restraint group depends on satisfaction of individual restraints in the group and the conditionality (all/any). A restraint group is considered satisfied, if the condition was met in at least one model of the model group/ensemble. The number of measured restraints can be smaller than the total number of restraint groups if crosslinks involve non-modeled residues. Only deposited models are used for validation right now.

State group	State	Model group	# of Deposited models/Total	Restraint group type	Satisfied (%)	Violated (%)	Count (Total=100)
1	1	1	1/1	All	Not available	Not available	0

#### Per-model satisfaction rates in ensembles

Every point represents one model in a model group/ensemble. Where possible, boxplots with quartile marks are also plotted.

### 5.3. 3DEM

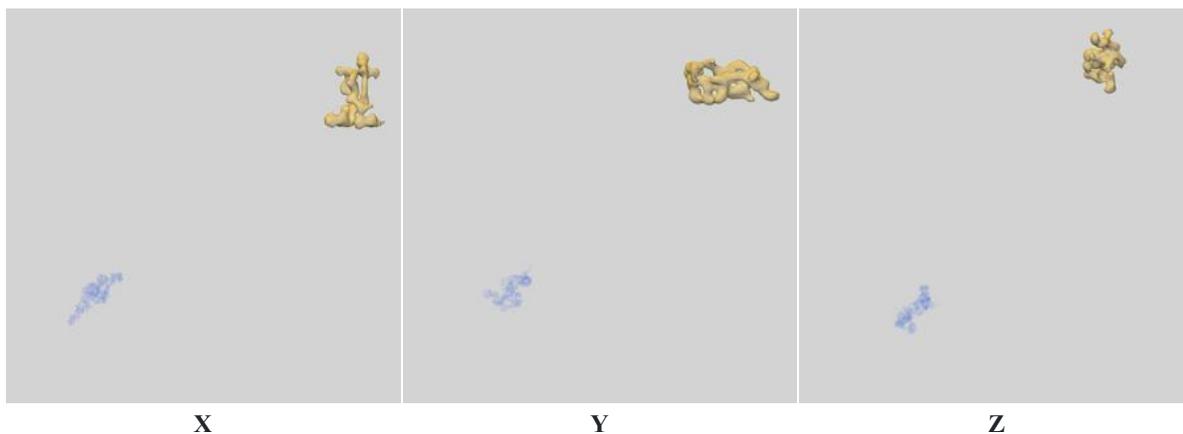
This section describes fit of models to the 3DEM data. Only results for the representative model, selected as a first model with the largest number of asymmetric units.

#### EMD-26791

##### 5.3.1. Map-model fit

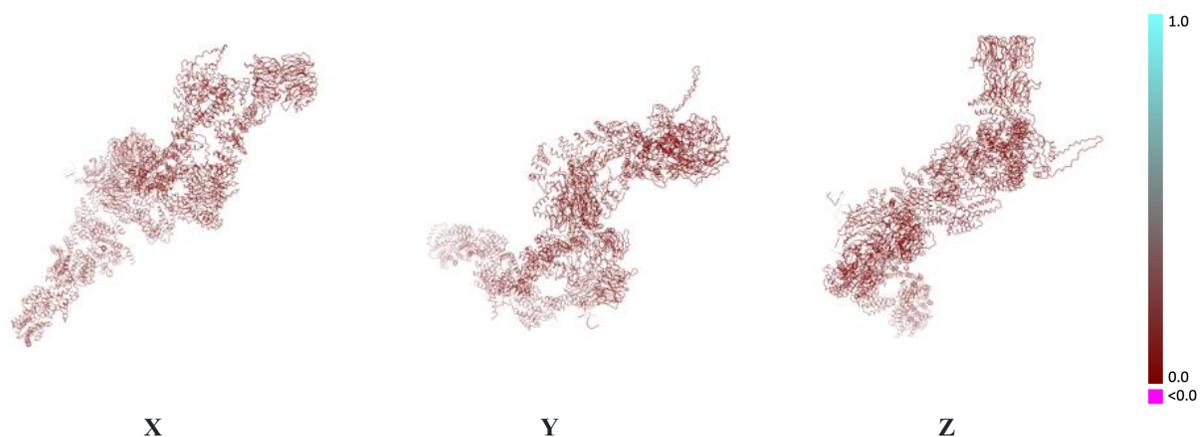
Only results for the representative Model 1 are shown.

##### 5.3.1.1 Map-model overlay



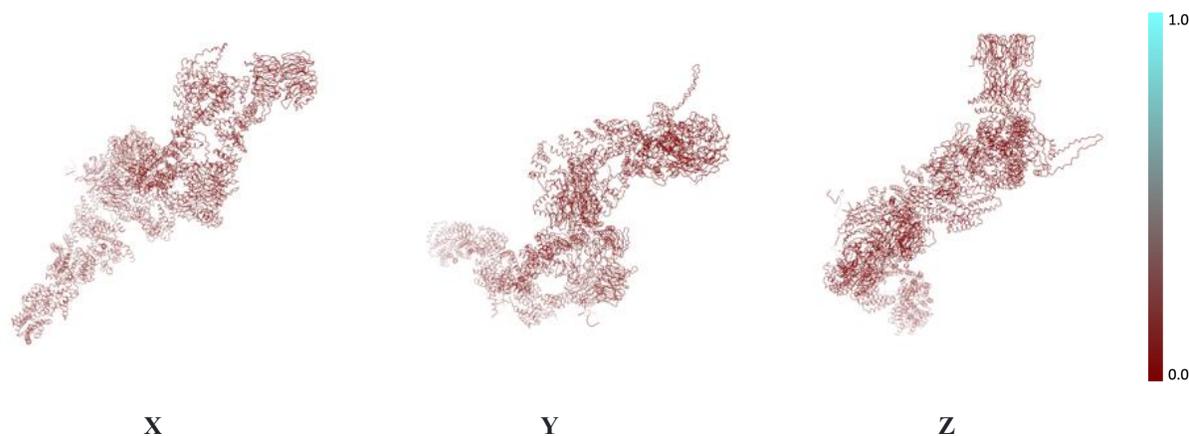
The images above show the 3D surface view of the map at the recommended contour level 1.150 at 50% transparency in yellow overlaid with a ribbon representation of the model colored in blue. These images allow for the visual assessment of the quality of fit between the atomic model and the map.

#### 5.3.1.2. Q-score mapped to coordinate model



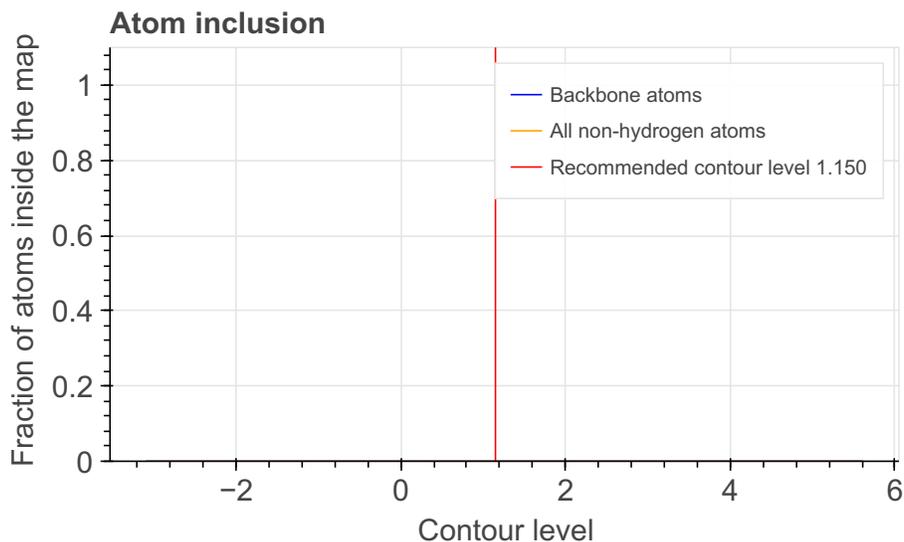
The images above show the model with each residue colored according to its Q-score. This shows their resolvability in the map with higher Q-score values reflecting better resolvability. Please note: Q-score is calculating the resolvability of atoms, and thus high values are only expected at resolutions at which atoms can be resolved. Low Q-score values may therefore be expected for many entries.

#### 5.3.1.3. Atom inclusion mapped to coordinate model



The images above show the model with each residue colored according to its atom inclusion. This shows to what extent they are inside the map at the recommended contour level 1.150 .

#### 5.3.1.4. Atom inclusion



At the recommended contour level, 0% of all backbone atoms, 0% of all non-hydrogen atoms, are inside the map.

#### 5.3.1.5. Map-model fit summary ?

The table lists the average atom inclusion at the recommended contour level ( 1.150 ) and Q-score for the entire model and for each chain.

Chain	Atom inclusion	Q-score
All	0.000	0.000
A	0.000	0.000
B	0.000	0.000
C	0.000	0.000
D	0.000	0.000
E	0.000	0.000
F	0.000	0.000

Color scale for Atom inclusion and Q-score: 1.0 (light blue), 0.0 (dark red), <0.0 (magenta).

## 6. Fit to Data Used for Validation Assessment ?

Validation for this section is under development.

### Acknowledgments

The development of integrative model validation metrics, implementation of a model validation pipeline, and creation of a validation report for integrative structures are funded by NSF awards to the *PDB-IHM team* (DBI-1756248, DBI-2112966, DBI-2112967, DBI-2112968, and DBI-1756250) and awards from NSF, NIH, and DOE to the *RCSB PDB* (DBI-2321666, R01GM157729, and DE-SC0019749). The PDB-IHM team and members of the *Sali lab* contributed model validation metrics and software packages.

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*crosslinking-MS validation methods. We are grateful to Dr. Shruthi Viswanath for discussions about uncertainty assessment of integrative structural models.*

*Members of the wwPDB Integrative/Hybrid Methods Task Force provided recommendations and community support for the project.*